An International Multidisciplinary e-Magazine www.sabujeema.com Volume 2 | Issue 11| November, 2022

# 

# **FISH HYDROLYSATE**

Jungivala Mansi Dipakbhai & Dr. B. G. Chudasama

# "Read More, Grow More"



Sabujeema Sabujeema f editorsabujeema@gmail.com sabujeema-international in multidisciplinary-e-magazine

31

and More, Grew



2



## FISH HYDROLYSATE [Article ID: SIMM0203]

### Jungivala Mansi Dipakbhai

M. F. Sc. (Fish Processing Technology), Dept. of Fish Processing Technology, College of Fisheries Science, Kamdhenu University

### Dr. B. G. Chudasama

Assistant Professor & Head, Dept. of Fish Processing Technology, College of Fisheries Science, Kamdhenu University

### ABSTRACT

ish protein hydrolysate (FPH) is resulting product of fish hydrolysate. FPHs are considered the most important source of protein and bioactive peptides and also possess excellent nutritional value since they have a balanced acid composition and a high amino digestibility. FPHs also possess bioactive properties such as anti-hypertension and anticancer activity. FPHs obtain through chemical hydrolysis, enzymatic hydrolysis or by fermented hydrolysis.

**KEYWORDS:** Fish protein hydrolysate, Hydrolysis, Enzymatic, Bioactive.

### **INTRODUCTION**

There is a rapid increase in the commercial seafood industry in the world in recent decades and fish processing units generating large quantities of solid waste and wastewater (Muzaddadi et al. 2016). According to Awarenet (2004), solid waste represents 20–60% of the initial raw material which consists of various kinds of residues, including whole waste fish, fish heads, viscera, skin, bones, blood, frame liver,

gonads, guts, and some muscle tissue, etc. The by-products, which are discarded, are used for plant fertilizer, livestock feed, and value-added specialty foods such as fish oil,which could be isolated efficiently and used as a dietary supplement (Ananey-Obiri et al. 2019). By-products are a significant source of protein, phospholipids, soluble vitamins, and bioactive compounds (Villamil et al. 2017). Fish peptides are shown anti oxidative, antihypertensive, and antithrombotic effects (Hamed et al. 2015).

Hydrolysates are the resulting products of hydrolysis (Ananey-Obiri et al. 2019). Schaafsma (2009) defined protein hydrolysates as a blend of short- and longchain peptides and amino acids that result from partial hydrolysis. Hydrolysates are considered the most important source of protein and bioactive peptides (Ananey-Obiri et al. 2019). FPH can be obtained by controlled enzymatic hydrolysis (Wang, 2021). FPHs possess excellent nutritional value since they have a balanced amino acid composition and a high digestibility (Wang, 2021). Fish protein hydrolysate (FPH) is a breakdown product of fish proteins that containing smaller peptides and amino acids (Venugopal, 2016).

### SOURCES OF FISH PROTEIN HYDROLYSATE

GTO FPH sources derive from three major fish components: muscle, skin, and waste (Wang, 2021). Fish muscles are rich sources of protein and account for 65%-75% (w/w) of the total protein which is used for extraction of FPH (Harnedy & Fitzgerald, 2013). Lean fish species or their processing wastes are the ideal raw material for FPH, which also can be used as food binders, emulsifiers, gelling agents, and nutritional supplements (Venugopal, 2016). Seafood proteases from

lore,



Volume 2 - Issue 11– November, 2022

An International Multidisciplinary e-Magazine



Atlantic salmon and trypsin from fish pyloric ceca have been used for FPH (Kristinsson & Rasco, 2000; Zamani & Benjakul, 2016).

### STRUCTURAL AND FUNCTIONAL PROPERTIES OF FISH PROTEIN HYDROLYSATE

Functional properties are defined as overall physiochemical properties of proteins in food systems during processing, storage, and consumption (Ananey-Obiri et al. 2019). Many functional properties of hydrolysates have been mentioned, such as solubility, emulsifying properties, foaming properties, water holding capacity, and fat binding capacity (Ananey-Obiri et al. 2019).

Solubility is the property that measuring how well the solute can dissolve in a solvent. A decrease in hydrogen bonds due to larger molecular weight causes reduced solubility. The hydrolysis process generates more carboxylic and amine groups, which provide more exposed groups to surrounding water (Ananey-Obiri et al. 2019).

The emulsifying properties of the hydrolysate are determined by the reduced tension of hydrophobic and hydrophilic components and a greater exposure to interaction at the oil-water interface. Hydroxylated amino acids contributed to the emulsion of hydrolysate from rainbow trout viscera (Taheri et al. 2013).

Foaming properties are governed by penetration and rearrangement at the airwater interface (Halim et al. 2016). Hydrophobic regions of the molecules determine the adsorption at the air-water interface (Ananey-Obiri et al. 2019).

Water holding capacity is the ability of the protein to capture water in the food matrix. Low molecular weight peptides are more hydrophilic and can affect water holding capacity more (Ananey-Obiri et al. 2019).

### BIOACTIVE PROPERTIES OF FISH PROTEIN HYDROLYSATE

Bioactive or biologically active peptides are beneficial for human health due to their positive effects on physiological activity such as that of anti-hypertension and anti-cancer activity (Wu et al. 2015). Marinederived peptides with strong antioxidant activity are able to scavenge free radicals and prevent lipid oxidation (Shavandi et al. 2019). During hydrolysis process, it generate peptides that exhibiting bioactivities such as anti-inflammation, antibacterial, or antioxidant activity (Chakrabarti et al. 2014). Other bioactive properties exist via hydrolysis and exhibit anti-oxidation, antiproliferation, cyto-immunomodulation, antihypertension, and anti-diabetic activities (Ananey-Obiri et al. 2019).

### PRODUCTION OF FISH PROTEIN HYDROLYSATE

Fish protein hydrolysates (FPH) are a product made from fish or fish material by the method of protein hydrolysation (Petrova et al. 2018). FPH is produced in two forms: liquid and dried. Liquid FPH is a watering mixture of hydrolyzed proteins, which consist up to 90% of moisture (Petrova et al. 2018). FPH in a liquid form is highly unstable for a long-term storage and, it is also difficult to be transported. Thus, dried FPH is preferable due to a longer shelf-life, easier storage and transportation (Petrova et al. 2018).

The hydrolysis of protein achieves through acids, alkalis or enzymes mediated breakdown of proteins in the waste into smaller protein fractions, peptides and free



### Volume 2 - Issue 11– November, 2022

amino acids (Kim & Dewapriya, 2015). Acid hydrolysis makes the product unpalatable because of tryptophan destruction and the formation of sodium chloride after the neutralization (Kim & Dewapriya, 2015). Alkaline hydrolysis produces some toxic compounds which are undesirable for human consumption (Kim & Dewapriya, 2015). Among protein hydrolyzing methods, enzymatic hydrolysis offers several advantages over others (Kristinsson & Rasco, tiona 2000).

### Chemical hydrolysis

Chemical hydrolysis is provided by either acid or alkali. It is achieved by cleaving the bonds between different peptide groups in the protein sequence by chemical agents (Petrova et al. 2018).

### 1. Acid hydrolysis

hydrolysis, either In acid hydrochloric acid or sulfuric acid is used to completely hydrolyze the fish proteins at high temperatures, and most times under high pressure (Kristinsson & Rasco, 2000). High temperature and pressure of solution (121-138 °C and 220-310 mPa correspondingly) are held during several hours (typically 2–8) so because of that achieve a certain degree of hydrolysis (DH) (Pasupuleti & Braun, 2008). Then the mixture is neutralized to pH 6.0-7.0and delivered for further dehydration (Kristinsson & Rasco, 2000). Hydrolysates recovered by using this method have high solubility, but they are bitter and have reduced nutritional qualities and poor functionality (Chobert et al. 1996). It is a lowcost, easy and simple process so because of that makes it important at the industrial level (Das et al. 2021). However during acid hydrolysis, important amino acids, such as tryptophan, methionine, & cystine are lost. In addition, asparagine and glutamine are

converted into aspartic acid and glutamic acid respectively (Das et al. 2021).

### 2. Alkaline hydrolysis

alkaline hydrolysis, sodium In hydroxide is mainly used (Das et al. 2021). Alkali hydrolysis is held at less elevated temperatures (typically 27-54 °C) in the presence of such alkaline agents as calcium, sodium or potassium hydroxide during several hours until the desired degree of hydrolysis is reached (Pasupuleti & Braun, 2008). A number of amino acids as serine and threonine are destroyed during alkaline hydrolysis (Pasupuleti & Braun, 2008). Alkaline hydrolysis is limitedly used in the fish processing industry to recover a range of proteins from fish material and fish protein concentrates (Kristinsson & Rasco, 2000).

### **Enzymatic hydrolysis**

Enzymatic hydrolysis also known as proteolytic hydrolysis, which is the most common method of protein hydrolysate recovery from fish processing by-products (Ananey-Obiri et al. 2019). This process is carried out by using endogenous enzymes or is an accelerated and controlled process using exogenously sourced enzymes (Ananey-Obiri et al. 2019). Enzymatic hydrolysis is a partial hydrolysis, and by this method the functionalities of native proteins are improved (Althouse et al. 1995).

Enzymatic hydrolysis is carried out from one to several hours at mild conditions: slightly elevated temperatures (generally around 35–65 °C) and a certain pH according to the optimal requirements of the used enzymatic systems: alkali, neutral or acid (Petrova et al. 2018). For FPH production, the used enzymes are of animal (pepsins), plant (papain, bromelain) or microbial origin (alcalase, neutrase, flavourzyme) (Petrova et



Volume 2 - Issue 11– November, 2022

al. 2018). The inactivation of enzymatic hydrolysis is typically accomplished by the elevated temperatures of around 75–100 °C applied for 5–30 min (Kristinsson & Rasco, 2000). This process involves a high production cost due to the long reaction time (Ananey-Obiri et al. 2019). However, the resulting FPHs are more marketable and valuable (Shahidi et al. 1995).

Ault1d

### **Fermented hydrolysis**

biochemically Fermentation uses microorganisms to break down fish proteins into peptides and amino acids (Ananey-Obiri et al. 2019). Different microorganisms have been identified and used to produce hydrolysates from fish protein (Ananey-Obiri et al. 2019). Due to the differences in microorganisms used in the culture for fermentation, the functionality of FPHs recovered by this method may vary (Daliri et al. 2017). Protein hydrolysates have been produced through fermentations using bacteria such as Enterococcus faecium NCIM5335 (Balakrishnan et al. 2011), and lactic acid bacteria (LAB) Pediococcus acidilactici NCIM5368 (Chakka et al. 2015). It is also interesting to note that fermentation of fish protein can help remove hyperallergic or anti-nutritional components that are found in the ingredients (e.g., trypsin inhibitors, glycinin,  $\beta$ -conglycinin, phytate) (Hou et al. 2017). This offers an advantage that is not obtained from other FPH recovery methods (Ananey-Obiri et al. 2019).

### APPLICATION OF FISH HYDROLYSATE

FPH is used in dietetic foods, such as soufflés, meringues, macaroni, or bread, and for the preparation of fish soup, fish paste, and shellfish analogues as flavoring compounds as a source of small peptides and amino acids (Muzaddadi et al. 2016). The major feed applications of FPH are as milk replacers for calves and weaning pigs and as proteins and attractants in fish feed (Gildberg, 1993). Chemical composition of fish protein hydrolysates is important in nutrition perspective of human health (Elavarasan, 2019). In general, required essential amino acids are abundant in FPH with richness in glutamic and aspartic acid content (Elavarasan, 2019). Fish protein hydrolysates are soluble in wide range of pH which is an ideal characteristic helps to use in wide range of products (Elavarasan, 2019).

Fish protein hydrolysates (FPHs) have been used in aquaculture feeds in order to enhance the growth and survival of fish (Elavarasan, 2019). The amino acid composition and the peptides present in hydrolysate are responsible for the improved growth immunological and status (Elavarasan, 2019). FPH is also being used as a source of protein in poultry feed formulation and in pet animal foods (Elavarasan, 2019). Other applications include FPH as a plant booster, ingredient in microbiological media and as a cryoprotectant in fish mince/surimi (Elavarasan, 2019).

### **CONCLUSION**

FPH offers many useful capabilities, which makes its applicability in food processing and even outside food processing desirable. The functionality, bioactivity, and nutritional value of FPH widen its scope of use. The choice of method used in the hydrolysis process prodigiously impacts the physicochemical, functional, and bioactive characteristics of the FPH, and consequently its use as a food ingredient.

### REFEENCES







- Althouse, P. J., Dinakar, P., & Kilara, A. (1995). Screening of proteolytic enzymes to enhance foaming of whey protein isolates. *Journal of food science*, 60(5), 1110-1112.
- Ananey-Obiri, D., Matthews, L. G., &Tahergorabi, R. (2019). Proteins from fish processing by-products. In *Proteins: Sustainable source, processing and applications* (pp. 163-191). Academic Press.
- Balakrishnan, B., Prasad, B., Rai, A. K., Velappan, S. P., Subbanna, M. N., & Narayan, B. (2011). In vitro antioxidant and antibacterial properties of hydrolysed proteins of delimed tannery fleshings: comparison of acid hydrolysis and fermentation methods. *Biodegradation*, 22(2), 287-295.
- Chakka, A. K., Elias, M., Jini, R., Sakhare, P.
  Z., & Bhaskar, N. (2015). In-vitro antioxidant and antibacterial properties of fermentatively and enzymatically prepared chicken liver protein hydrolysates. *Journal of Food Science and Technology*, 52(12), 8059-8067.
- Chakrabarti, S. K., Singh, B. P., Thakur, G., Tiwari, J. K., Kaushik, S. K., Sharma, S., & Bhardwaj, V. (2014). QTL Analysis of late blight resistance in a diploid potato family of *Solanum spegazzinii× S. chacoense. Potato research*, 57(1), 1-11.
- Chobert, J. M., Briand, L., Guéguen, J., Popineau, Y., Larré, C., & Haertlé, T. (1996). Recent advances in enzymatic modifications of food proteins for improving their functional properties. *Food/Nahrung*, 40(4), 177-182.



- Daliri, E. B. M., Lee, B. H., & Oh, D. H. (2017). Current perspectives on antihypertensive probiotics. *Probiotics and antimicrobial proteins*, 9(2), 91-101.
- Das, A., Nayak, Y., & Dash, S. (2021). Fish protein hydrolysate production, treatment methods and current potential uses: A review. *Skin*, *8*, 12.
- Elavarasan, K. (2019). Health Benefits and Potential Applications of Fish Protein Hydrolysate. ICAR-Central Institute of Fisheries Technology, 65-78
- Gildberg, A. (1993). Enzymic processing of marine raw materials. *Process Biochemistry*, 28(1), 1-15.
- Halim, N. R. A., Yusof, H. M., & Sarbon, N.
  M. (2016). Functional and bioactive properties of fish protein hydolysates and peptides: A comprehensive review. *Trends in Food Science & Technology*, *51*, 24-33.
- Hamed, I., Özogul, F., Özogul, Y., & Regenstein, J. M. (2015). Marine bioactive compounds and their health benefits: a review. *Comprehensive reviews in food science and food safety*, 14(4), 446-465.
- Harnedy, P. A., & Fitzgerald, R. J. (2013).
  Bioactive proteins and peptides from macroalgae, fish, shellfish and marine processing waste. *Marine proteins and peptides: Biological activities and applications*, 5-39.
- Hou, Y., Wu, Z., Dai, Z., Wang, G., & Wu,
  G. (2017). Protein hydrolysates in animal nutrition: Industrial production, bioactive peptides, and functional significance. *Journal of Animal Science and Biotechnology*, 8(1), 1-13.







- Kim, S., & Dewapriya, P. (2015). Biomedical potential of fish processing byproducts. In N. M. Sachindra, & N. S. Mahendrakar (Eds.), Fish processing byproducts: quality assessment & applications (pp. 63-76). Studium Press LLC.
- Kristinsson, H. G., &Rasco, B. A. (2000). Fish protein hydrolysates: production, biochemical, and functional properties. Critical reviews in food science and nutrition, 40(1), 43-81.
- Muzaddadi, A. U., Devatkal, S., & Oberoi, H. S. (2016). Seafood enzymes and their application in food processing. In Agro-Industrial Wastes as Feedstock for Enzyme Production (pp. 201-232). Academic Press.
- Pasupuleti, V. K., & Braun, S. (2008). State of the art manufacturing of protein hydrolysates. Protein hydrolysates in biotechnology, 11-32.
- Petrova, I., Tolstorebrov, I., & Eikevik, T. M. (2018). Production of fish protein hydrolysates step by step: technological aspects, equipment used, major energy costs and methods of their minimizing. International Aquatic Research, 10(3), 223-241.
- Schaafsma, G. (2009). Safety of protein hydrolysates, fractions thereof and bioactive peptides in human nutrition. European journal of clinical nutrition, 63(10), 1161-1168.
- Shahidi, F., Han, X. Q., & Synowiecki, J. (1995). Production and characteristics of protein hydrolysates from capelin villosus). Food (Mallotus chemistry, 53(3), 285-293.
- Shavandi, A., Hou, Y., Carne, A., McConnell, M., & Bekhit, A. E. D. A.

(2019). Marine waste utilization as a of functional and source health compounds. Advances in food and nutrition research, 87, 187-254.

- Taheri, A., Anvar, S. A. A., Ahari, H., & Fogliano, V. (2013). Comparison the functional properties of protein hydrolysates from poultry by-products and rainbow trout (Onchorhynchus mykiss) viscera. Iranian Journal of Fisheries Sciences, 12(1), 154-169.
- V. (2016). Enzymes from Venugopal, seafood processing waste and their applications in seafood processing. Advances in food and nutrition research, 78, 47-69.
- Villamil, O., Váquiro, H., &Solanilla, J. F. (2017). Fish viscera protein hydrolysates: Production, potential applications and functional and bioactive properties. Food Chemistry, 224, 160-171.
- Wang, X. (2021). Natural bioactive compounds from fish. Natural Bioactive *Compounds*, 393-408.
- Wu, R., Wu, C., Liu, D., Yang, X., Huang, J., Zhang, J., ... & Li, H. (2015). Overview of antioxidant peptides derived from marine resources: The sources, characteristic, purification, and evaluation methods. Applied biochemistry and biotechnology, 176(7), 1815-1833.
- Zamani, A., &Benjakul, S. (2016). Trypsin from unicorn leatherjacket (Aluterus monoceros) pyloric caeca: purification and its use for preparation of fish protein hydrolysate with antioxidative activity. Journal of the Science of Food and Agriculture, 96(3), 962-969.